International Journal of Novel Research in Physics Chemistry & Mathematics Vol. 10, Issue 3, pp: (63-86), Month: September - December 2023, Available at: <u>www.noveltyjournals.com</u>

SOLID STATE FERMENTATION OF RICE BRAN: NUTRITIONAL VALUES AND FUNCTIONAL PROPERTIES

¹Shehu Isah, ²Kennedy Unakalamba

¹Caleb University, Lagos and ²University of Abuja, Abuja DOI: <u>https://doi.org/10.5281/zenodo.8366646</u> Published Date: 21-September-2023

Abstract: Solid state fermentation of rice bran improves nutritional values and functional properties. The edible fungus, *Pleurotus Sapidus*, was employed for the solid state fermentation. During fermentation, the sample was withdrawn after ten days and further analyzed. An investigation process was carried out on the solid-state fermented rice bran (RB) in comparison with the unfermented (normal) rice bran. The few analyses that were investigated in comparison with the unfermented rice bran were the density tests (bulk, tapped and compact), water and oil absorption capacities, swelling power, moisture estimate, pH, reducible sugar, and water solubility. From the results, it was found that both the fermented and unfermented rice bran have a pH of 5 and 6, respectively, with absorption capacities of both oil (5% for the fermented sample and 20% for the unfermented sample) and water (10% for the fermented sample and 25% for the unfermented sample), and both samples possess swelling power. This study demonstrated a comparison in the nutritional quality of RB after fermentation with *Pleurotus sapidus* in an attempt to find or improve the functional and nutritional value of rice bran via solid state fermentation. The study therefore proves that the functional and nutritional value of the unfermented sample, which has a higher phenol concentration from spectrophotometry and high ascorbic acid, was better than the fermented sample with lower phenol concentration and less ascorbic acid.

Keywords: Solid state fermentation, rice bran, high ascorbic acid.

1. INTRODUCTION

Rice bran oil, containing 90-96% lipid, is considered a healthy food due to its high mono- and polyunsaturated fatty acids content. Within the fatty acid (FA) fraction, palmitic acid (21-26%), linoleic acid (31-33%), and oleic acid (37-42%) are the predominant compounds [1-10]. However, its short shelf life is due to its high lipid percentage and lipase enzymes, which degrade the oil, making it rancid and unsuitable for consumption.

Rice bran and its oil are used for medicinal purposes, particularly in Japan, Asia, and India. They are used for conditions like diabetes, high blood pressure, and cholesterol. Rice bran oil contains substances like Vitamin E, which may help lower cholesterol and reduce kidney stone formation. However, there is limited scientific evidence supporting these claims [12].

BASIC COMPOSITION OF RICE BRAN

It's apparent that rice bran is a potential source of high-value antioxidants for use as additives in foods, pharmaceuticals, and cosmetics because of its unsaturated fatty acid as synergists for antioxidants ^{[66]; [67]; [68]}.

Earlier studies showed rice bran compositional distinctiveness such as carbohydrate, protein, fat, moisture, ash, fiber, amylose contents, phytic acid, minerals, vitamin contents, and the Glycemic Index (GI) ^{[69]; [70]}. Previous analysis of rice

Vol. 10, Issue 3, pp: (63-86), Month: September - December 2023, Available at: www.noveltyjournals.com

varieties in a particular region of Bangladesh and found they are composed of protein (7.04%), fat (0.37%), crude fiber (0.26%), and ash (0.58%) in parboiled milled rice ^[71].

Moreover, aromatic rice varieties showed exciting composition such as moisture (11.25%-15.13%), protein (3.23%-6.21%), fat (0.68%-1.45%), and ash (0.88%-1.46%) ^[72].

The functional composition of rice bran contains a rich source of bioactive compounds ^[29]. Rice bran's health benefits and enhanced quality have been reported due to their antioxidant compounds and health benefit. It's apparent that rice bran is a potential source of high-value antioxidants for use as additives in foods, pharmaceuticals, and cosmetics because of its unsaturated fatty acid as synergists for antioxidants ^[66]; ^[67]; ^[68].

A recent study has quantified the functional compounds in rice bran including γ -oryzanols (mixture of lipids derived from rice which occurs mainly in the fat fraction of rice bran and rice bran oil) and vitamin E components (tocopherols) in rice bran oil ^[73], anthocyanin components in red rice ^[74], and phenolic acids in various rice varieties ^[75].

SUBMERGED LIQUID FERMENTATION (SLF)

Submerged liquid fermentation (SLF) is a method used in industrial manufacturing to produce bio-molecules by submerging enzymes and reactive compounds in liquids like alcohol, oil, or nutrient broth. This process requires liquid media for microorganism growth, but higher water content can decrease concentration and contaminate media. The higher quantity of liquid waste produced leads to dumping issues. SLF is expensive due to the need for processed substrate and large-scale practice, increasing labour costs [13-16].

Solid state fermentation (SFF) is a method used to enhance oil recovery from shredded coconut meat, particularly in Indonesia. It is increasingly being used over submerged liquid fermentation due to its low cost, lack of liquid for media preparation, and lower energy consumption, plant and machinery costs, and labor costs [17]. SSF produces high product yields with higher stability and productivity, but it also has disadvantages like delayed enzyme production and toxic substances. Biotechnological industries face challenges in end product purification, scale-up, and biomass estimation [18]. Rice bran fermentation has been studied for its functional properties, revealing increased nutrient availability, bio-surfactant content, and fatty acid content. This process also allows the release and transformation of phenolic and volatile compounds, further enhancing the potential benefits of fermented substrates [19].

An array of health-promoting value added products have been derived from processed rice bran due to its identified active components such as oryzanols, tocopherols, tocotrienols, phytosterols, nucleotides, dietary fiber content, and phenolic compounds ^{[33]; [34]; [35]}.

The three main constituent of include; cycloartenylferulateoryzanol A, 24-methlenecycloartanyl ferulateoryzanol C and campesterylferulate, as shown in fig 3, 4 and 5 below.

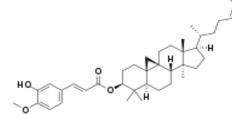


Fig 3: Cycloartenylferulateoryzanol A (C₄₀H₅₈O₄)

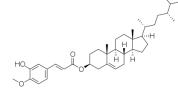


Fig 5: Campesterylferulate (C₃₈H ₅₆O₄)

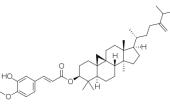


Fig 4: 24-methylenecycloartanyl

Vol. 10, Issue 3, pp: (63-86), Month: September - December 2023, Available at: www.noveltyjournals.com

As shown in fig. 6 below, both tocopherols and tocotrienols occur in α (alpha), β (beta), γ (gamma) and δ (delta) forms, determined by the number and position of methyl groups on the chromanol ring.

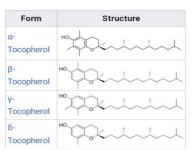
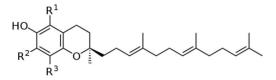


Fig 6: The different forms of tocophenols



General chemical structure of tocotrienols. alpha(a)-Tocotrienol: R1 = Me, R2 = Me, R3 = Me; $beta(\beta)$ -Tocotrienol: R1 = Me, R2 = H, R3= Me; $gamma(\gamma)$ -Tocotrienol: R1 = H, R2 = Me, R3= Me; $delta(\delta)$ -Tocotrienol: R1 = H, R2 = H, R3= Me

Fig 7: Tocotrienol, showing how the different forms can be achieved

Phytosterols (plant sterol) are a group of naturally occurring compounds found in plant cell membranes, with structure similar to cholesterol. There are of two groups; plant sterols (those with double bonds in the sterol rings) and plant stanols (those with no double bonds in the sterol rings)

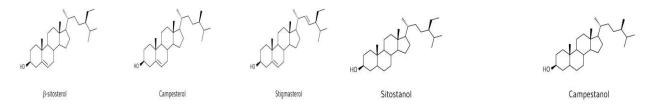


Fig 8: Plant sterols diagram

Fig 9: Plant stanols diagram

Coloured or pigmented rice (purple, black, and red rice) is one of the major food items in an Asian-based diet ^[38]. The constituents of coloured rice are flavonoids, phenolics, tannin, sterols, tocols, γ -oryzanols, amino acids, fatty acids, phytoantioxidant compounds, vitamins, and dietary fibers^{[39]; [40]; [41]}. Although rice bran usage is limited due to its rapid rancidity and unfavourable aroma, rice bran fermented with different types of microorganisms makes it an effective agent to retain its potential therapeutic efficacy.

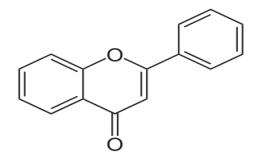


Fig. 11: Flavone, an example of flavonoids

Vol. 10, Issue 3, pp: (63-86), Month: September - December 2023, Available at: www.noveltyjournals.com

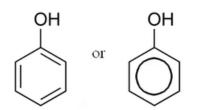


Fig. 12: Skeletal structure of phenolic compound

Gallic acid, Phloroglucinol and flavan-3-ol's scaffold are the three base units or monomer of tannin and their classes/polymer are hydrolysable, phlorotannins and condensed tannins and phlobatannins (C-ring isomerized condensed tannins) respectively.

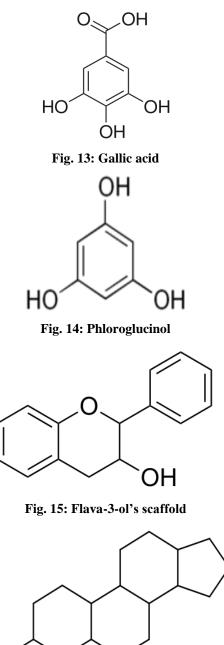


Fig. 16: Sterols

HO

Vol. 10, Issue 3, pp: (63-86), Month: September - December 2023, Available at: www.noveltyjournals.com

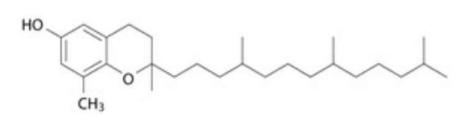


Fig. 17: Tocols

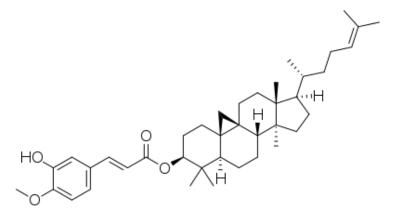


Fig. 18: γ-oryzanol (Cycloartenylferulateoryzanol A)

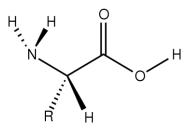


Fig. 19: L-amino acid (an example of amino acid)

The main phyto-antioxidants compounds are polyphenols and carotenoids and their structures are shown below.

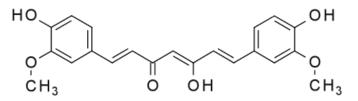


Fig. 21: Curcumin (an example of polyphenol)

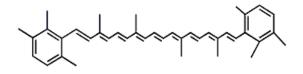


Fig. 22: Isorenieratene (C40H48) (General structure of carotenoid)

Vol. 10, Issue 3, pp: (63-86), Month: September - December 2023, Available at: www.noveltyjournals.com

RICE BRAN EFFECT ON INFLAMMATORY BOWEL DISEASE (IBD)

Fermented rice bran (FRB) is a bioactive compound from rice bran that has been shown to have protective effects against various diseases, including cancer, metabolic syndrome, obesity, diabetes, and immune modulation. However, there is limited research on FRB's potential against inflammation-related diseases like inflammatory bowel disease (IBD) and multifactorial metabolic disorder (MMD). IBD is a chronic inflammation in the gastrointestinal tract, linked to an increased risk of colon cancer and colorectal cancer. Urbanization, diet, antibiotic use, hygiene, microbial exposures, and pollution are also potential risk factors for IBD and MMD. Injuries in the intestinal mucosa damage the barrier function, leading to inflammation and the production of excessive pro-inflammatory cytokines [42-47].

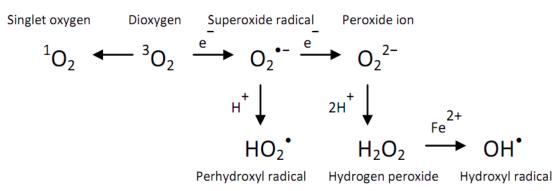


Fig. 23: Illustration of oxidative stress

Rice bran has been found to have anti-diabetic and anti-dyslipidemic properties, potentially aiding in treating multifactorial metabolic disorder and colonic disorders. Fermented rice bran with Saccharomyces cerevisiae and Lentinusedodes has antistress, anti-fatigue, anti-cancer, and anti-defective immune responses. Ferulic acid and phenolic compounds have hypoglycemic effects in type 2 diabetic mice, and Driselase-treated rice bran fraction improves glucose and lipid metabolism in SHRSP rats [47,49,51-53].

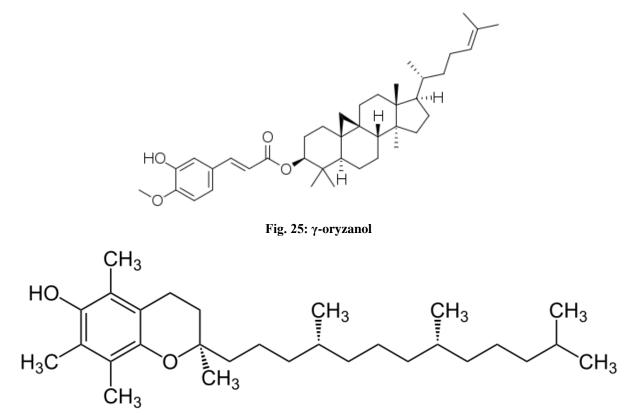


Fig. 26: Alpha-Tocopherol (Vitamine E component)

Vol. 10, Issue 3, pp: (63-86), Month: September - December 2023, Available at: www.noveltyjournals.com

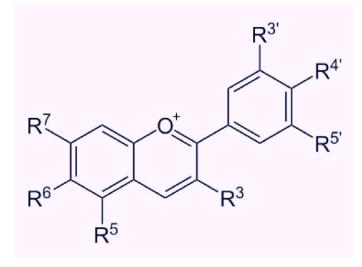


Fig. 27: Anthocyanin

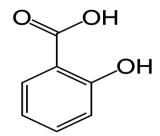


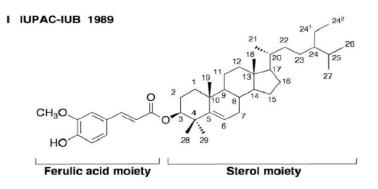
Fig. 28: Salicylic acid (Phenolic acid)

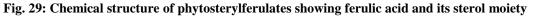
RICE BRAN-BASED FUNCTIONAL FOOD, A DRUG ALTERNATIVE

Functional food is food products fortified with special constituents with advantageous physiological effects. It can provide health benefits beyond traditional nutrients, as defined by the 1994 National Academy of Sciences' Food and Nutrition Board definition. Functional foods can improve health conditions and homeostatic behavior, using biomarkers or indicators in body homeostasis. Rice bran foods have been found to have a positive effect on human health, with antihypertensive effects.[76-82]

PREVENTIVE ROLE OF FERMENTED RICE BRAN ON TUMORIGENESIS

The study by Kuno et al (2000) found that fermented brown rice and rice bran with A. oryzae (FBRA) can prevent prostate tumorigenesis in transgenic rats. The supplementation decreased adenocarcinoma in the lateral prostate, suppressed prostate carcinogenesis, increased apoptosis, and inhibited cell proliferation in high-grade prostatic intraepithelial neoplasias. It also limited tumor growth by activating energy deprivation pathways, indicating its potential to prevent human prostate cancer [58,83]





Vol. 10, Issue 3, pp: (63-86), Month: September - December 2023, Available at: www.noveltyjournals.com

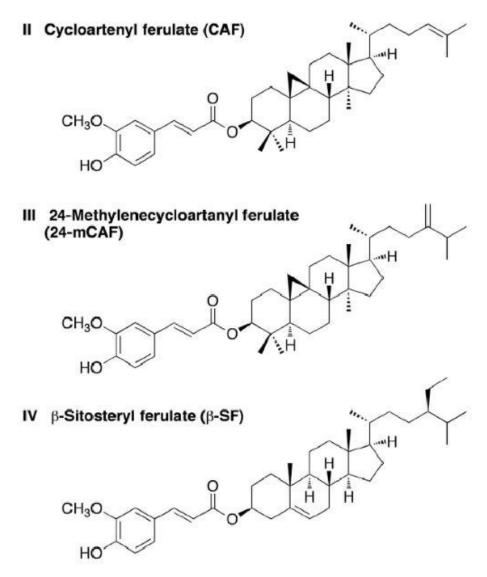


Fig. 30: Some examples of phytosterylferulate: cycloartenylferulate (CAF, principal components of oryzanol), 24methylenecycloartanyl ferulate (24-mCAF), and sitosterylferulate (SF)

SWELLING CAPACITY (SWELLING INDEX)

The swelling index (SI), also known as swelling capacity (SC), is the volume taken up by one gram of food material under specific conditions. It measures starch's ability to absorb water and swell, reflecting associative forces in granules. Swelling capacity is a quality measure in some food products, such as bakery ones, and is influenced by particle size, species variety, and processing methods[10,100]

OIL ABSORPTION CAPACITY (OIL ABSORPTION)

Oil absorption capacity (OAC) is the binding of fat by non-polar side chains of proteins, which enhances mouth feel and retains food products' flavor. It is highly prevalent in foods with high protein content and depends on factors like protein conformation, amino acid composition, and surface polarity. Food substances with good OAC capacities are useful in food applications where optimal oil absorption is desired, such as pastries production. Oil absorption during frying is a concern, as it increases the caloric value of food products. The ideal frying temperature is between 162.78°C (325°F) to 190.56°C (375°F). High OAC food substances are beneficial in structural interactions, improving palatability, extending shelf life, and flavor retention, especially in meat or bakery products. Protein, composed of both hydrophobic and hydrophilic parts, affects OAC capacity[100-103].

Vol. 10, Issue 3, pp: (63-86), Month: September - December 2023, Available at: www.noveltyjournals.com

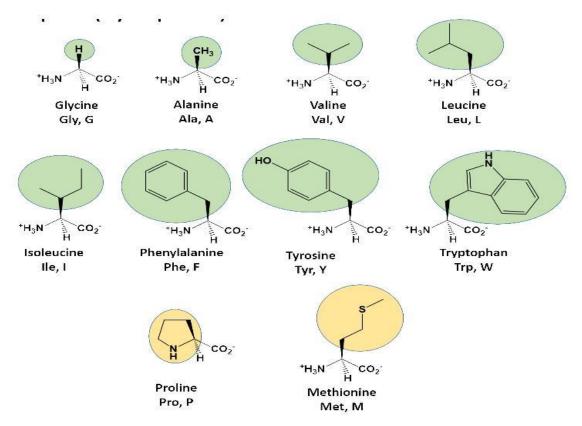
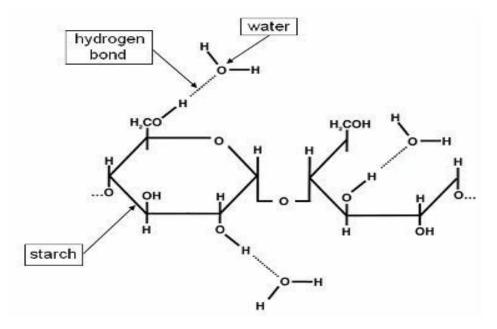
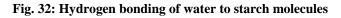


Fig. 31: Binding of oil with protein amino acid

WATER ABSORPTION CAPACITY (ALSO KNOWN AS WATER HYDRATION OR WATER ABSORPTION)

Water absorption capacity (WAC) is the amount of water taken up by a substance to achieve desired consistency and quality food products. It is often defined by the weight of the substance. Water absorption occurs when water and food substance mix, hydrating gluten-forming proteins, damaged starch, and other ingredients through hydrophilic interactions and hydrogen bonds with water molecules.





Vol. 10, Issue 3, pp: (63-86), Month: September - December 2023, Available at: www.noveltyjournals.com

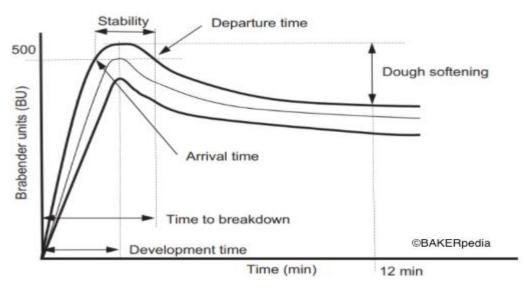


Fig. 33: Typical farinograph curve

Factors influencing the water absorption capacity of a food substance include; starch, protein, pentosans, vital wheat gluten (VWG), and presence of other water binding ingredients such as fiber, bran hydrocolloids (gums).

Water absorption capacity in foods is influenced by hydrophilic components like carbohydrates, proteins, and polar amino acids. Lower absorption in some flours may be due to less availability of polar amino acids. Increased water absorption may also be due to amylose solubility, leaching, and loss of starch structure. High water absorption capacity in composite flours suggests that different flours can be used in food formulations like processed cheese, bakery products, sausage, and dough. Water absorption capacity is crucial for consistency, bulking, and baking applications[100,108-110]

BULK DENSITY (ALSO KNOWN AS VOLUMETRIC DENSITY OR APPARENT DENSITY)

Bulk density is the mass of many flour particles divided by their total volume. It is a functional property of flours, powders, fine particles, granules, and other divided solids of foods. It can change depending on handling and can be improved by factors such as geometry, measurement method, particle size, surface properties, and solid density. Bulk density also influences the porosity of a food product, which impacts the design of the package and can determine the type of packaging material. Recent studies suggest that flour bulk density may be influenced by the initial moisture content, with high bulk density suggesting suitability for food preparations and low density for complementary foods. Starch forms the main structure and bulk of many food products[101,100].

SOLUBILITY

Solubility is the ability of food substances to dissolve in a solvent, typically water or oil. It depends on the chemical and physical properties of the solvent and solute, as well as pressure, pH, temperature, and other chemicals. Solubility is measured as the saturation concentration, where more solute does not increase the solution's concentration but precipitates excess solute. Lipids can reduce water absorption capacity, affecting solubility. High solubility indicates high digestibility, making it suitable for infant formula and food. Insolubility refers to the inability of a food to dissolve in a solvent. Water is the most common solvent in food, containing various organic, inorganic, and ionic compounds.[114]

2. MATERIALS AND METHODS

STATE FERMENTATION

10g of Potato Dextrose Agar (PDA) was measured and mixed with 250ml of water. 9ml was measured and kept separately to mix with the sample. The potato dextrose agar solution was then sterilized with the 9ml of water that was measured out separately, in an autoclave machine at 121°C for 15mins. After sterilization, they were allowed to cool for a while and then 10g of the sample (Rice bran) was mixed with the 9ml of sterilized water. The sample solution was poured into the sterilized potato dextrose agar solution and stirred very well. The solution was poured into Petri dishes and labelled, wrapped with

Vol. 10, Issue 3, pp: (63-86), Month: September - December 2023, Available at: www.noveltyjournals.com

foil paper tightly to avoid reaction with free bacteria in the air, and incubated at 25°C and observed for 10 days. The sample that was gotten after 10 days was then used for further analysis in comparison with the normal, dry unfermented sample.



Fig. 51: Fermented sample (After 10 days)

BULK DENSITY (pB)

3g of sample, fermented and unfermented, was measured separately and poured into a measuring cylinder, and then the volume was observed and recorded. The mass of the sample (3g) was divided by the volume, which gave the bulk density.

TAPPED DENSITY (PT)

3g of sample, fermented and unfermented, was measured separately and poured into a measuring cylinder, "tapped severally" (in this experiment, it was tapped 6 times), and then the volume was observed and recorded. The mass of the sample (3g) was divided by the tapped volume, which gave the tapped density.

COMPACT DENSITY

5g of sample, fermented and unfermented, was measured separately and poured into a measuring cylinder, "compressed" (like pounded) until no further reduction in volume, and then the volume was observed and recorded. The mass of the sample (5g) was divided by the compressed volume, which gave the compact density.

CARR INDEX (C)

$$C = 100 \left(1 - \frac{pB}{pT}\right)$$

Where C = Carr index, pB = Bulk density and pT = Tapped density

The result from the tapped density (pT) and bulk density (pB) was used to calculate the Carr index, using the formula above.

HAUSNER'S RATIO (H)

$$H = pT/pB$$

The result from the tapped density (pT) and bulk density (pB) was also used to calculate the Hausner's ratio, using the formula above.

WATER ABSORPTION CAPACITY

10ml of distilled water was mixed with 1g of sample (both samples, separately) and allowed to settle for 30mins after which it was centrifuged at 2000rpm for 10mins. After centrifuge, the liquid was separated from the solid and the weight and volume was observed and recorded.

Vol. 10, Issue 3, pp: (63-86), Month: September - December 2023, Available at: www.noveltyjournals.com

OIL ABSORPTION CAPACITY

10ml of groundnut oil was mixed with 1g of sample (both samples, separately) and allowed to settle for 30mins after which it was centrifuged at 2000rpm for 10mins. After centrifuge, the liquid was separated from the solid and the weight and volume was observed and recorded.

SWELLING POWER

14ml of distilled water was mixed with 0.3g of sample (both samples, separately) and heated up to 73°c and allowed to cool at room temperature for a while. After cooling, it was centrifuged at 5000rpm for 10mins. The fermented and unfermented sample was separated from the water after centrifuge, and the weight of both samples were recorded.

ESTIMATION OF MOISTURE

Estimated moisture = (lost weight / initial weight) \times 100

3g of both samples were measured separately and put in petri dishes and then put in an oven to dry at 105°C. A check was done at interval of 30mins until constant weight was obtained.

PH TEST

1g of both samples was mixed with 10ml of distilled water separately and then tested with litmus paper and the visible colour change of the litmus paper was observed.

REDUCIBLE SUGAR

1g of sample was mixed with 10ml of distilled water separately, and then 5 drops of Benedict's reagent (sodium carbonate, sodium citrate and copper (II) sulphate pentahydrate) was added to both separate mixture of samples and then heated up to 75° c and observed for any colour change which will indicate the presence of reducible sugar.

WATER SOLUBILITY

0.3g of both samples were added to 0.5ml of distilled water and then tapped to observe if any particle goes down. A solution of the sample was also prepared separately, and then added in water in drops and observed.

SPECTROPHOTOMETRIC METHOD FOR PHENOLIC COMPOUND (y-ORYZANOL) DETERMINATION

A mixture of 4ml of phenol was added to 1g of fermented and unfermented samples, mixed with ferric chloride, distilled water, and distilled water. The mixture was then diluted with distilled water and tested using a UV spectrophotometric machine. The concentration of phenol was determined using Beer Lambert's principle of absorbance, which was then treated with the molar attenuation coefficient of γ -oryzanol, a phenolic compound in rice bran.

ASCORBIC ACID

A little light blue solution of dichlorophenolindopheol (DCPIP), already diluted, was put in a 2 beakers, separately, for the 2 samples. Both samples solutions (fermented and unfermented) were prepared also. With the use of a pipette, a clear solution of the samples was drawn and then added to the dichlorophenolindophenol solution in the beaker, 1 drop at a time, to observe for colour change (blue to red or pink), which signifies the presence of ascorbic acid.

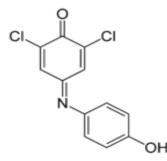


Fig. 52: (DCPIP) 2,6-dichlorophenolindophenol (C12H7Cl2NO2)

Vol. 10, Issue 3, pp: (63-86), Month: September - December 2023, Available at: www.noveltyjournals.com

3. RESULTS AND DISCUSSION

From the analysis carried out, table 1 below is the result, showing the bulk density, tapped density, compact density, Carr index and Hausner's ratio of the fermented and unfermented sample.

TESTS	FERMENTED SAMPLE	UNFERMENTED SAMPLE
Bulk Density	0.3g/ml	0.286g/ml
Tapped Density	0.75g/ml	0.3g/ml
Compact Density	0.833g/ml	0.385g/ml
Carr Index	60	4.67
Hausner's Ratio	2.5	1.05

Table 1: Density tests, Carr index and Hausner's ratio

BULK, TAPPED, AND COMPACTED DENSITY

These are functional properties of powdery substances or particles which can be used to determine the starch content in food particles and the higher the starch content, the more likely the increase in these densities. More also when there is an increase in the volume, then there will be a decrease in the bulk, tapped and compacted density, which indicates lower starch content. High bulk density also suggests suitability for application in food preparation. On the other hand, low density can also be useful in the formulation of complementary foods.

So from the result above, the fermented sample obviously has a higher bulk, tapped and compacted density therefore contains more starch content. In other words, fermented sample is more suitable for application of food preparation. Meanwhile, on the other hand, the unfermented sample with low density would be more useful in the formation of complementary foods.

CARR INDEX

Carr index is an indication of the compressibility of a powder and is frequently used in pharmaceutics. In a free-flowing powder, the bulk density and tapped density would be close in value; therefore, the Carr index would be small. On the other hand, in a poor-flowing powder where there are greater inter-particle interactions, the difference between the bulk and tapped density observed would be greater, therefore, the Carr index would be larger ^[147]. A Carr index greater than 25 is considered to be an indication of poor flowability, and below 15, of good flowability^[148].

The bulk and tapped density of the fermented sample is not close, from the table above, and obviously, the carr index is very large (greater than 25), therefore it is poor-flowing and has greater inter-particle interaction. On the other hand, the bulk and tapped density of the unfermented sample is very close, and the Carr index from the table is very low (below 15), so it is a good-flowing powder.

HAUSNER'S RATIO

Hausner's ratio is an indication of the flowability of a powder $^{[155]}$, and it's much related to Carr index. A Hausner ratio greater than 1.25 - 1.4^[156] is considered to be an indication of poor flowability.

The table above shows the Hauser's ratio of both the fermented and unfermented sample, after due calculation. The fermented sample can be considered to have poor flowability because it has a ratio above 1.25 - 1.4.

Other tests;

Table 2 below, shows the final results for the water/oil absorption capacity, swelling power, estimated moisture content, pH and the phenolic compound determination via spectrophotometry for both the fermented and unfermented sample.

 Table 2: Water/oil absorption capacity, swelling power, estimation of moisture, pH and spectrophotometric determination of phenolic compound (γ-oryzanol)

TEST	FERMENTED SAMPLE	UNFERMENTED SAMPLE
Water Absorption Capacity	10%	25%
Oil Absorption Capacity	5%	20%
Swelling Power	0.1g	0.8g

Vol. 10, Issue 3, pp: (63-86), Month: September - December 2023, Available at: www.noveltyjournals.com

Estimation of Moisture	97%	12.3%
Ph	5	6
Spectrophotometric Determination of Phenolic		
Compound (<i>y</i> -Oryzanol)	0.0105m/cm	2.105m/cm

ESTIMATION OF MOISTURE

The table below (Table 3) shows the different weights of 3g of both samples each, in the oven at interval of 30mins check.

	Initial weight	After 30mins	After 1hr	After 1:30mins	After 2hrs	After 2:30mins	Weight Lost
Fermented Sample	3g	1.56g	0.18g	0.11g	0.09g	0.09g	2.91g
Unfermented Sample	3g	2.64g	2.63g	2.63g	2.63g	2.63g	0.37g

 Table 3: Different weights at 30mins interval

SPECTROPHOTOMETRIC DETERMINATION OF PHENOLIC COMPOUND (7-ORYZANOL)

Spectrophotometric determination involves wavelength and infrared rays passing through a photometric machine to determine γ -oryzanol concentration. This knowledge is crucial in pharmaceuticals, especially for determining medication dosage. γ -oryzanol is used in Japan for menopausal symptoms, anxiety, stomach upset, and high cholesterol. Fermented samples have lower concentrations, while unfermented samples are richer.[158]

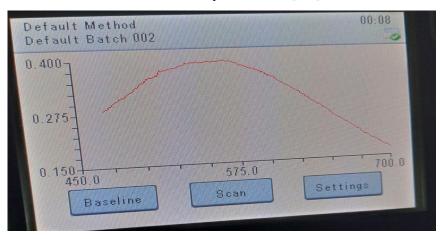


Fig. 58: Spectrophotometric phenol test for fermented sample

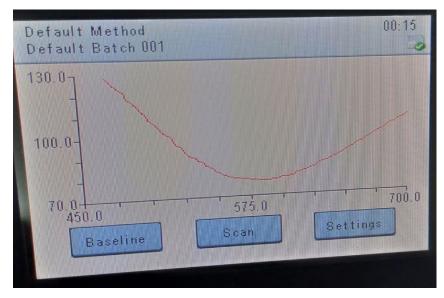


Fig. 59: Spectrophotometric phenol test for unfermented sample

Vol. 10, Issue 3, pp: (63-86), Month: September - December 2023, Available at: www.noveltyjournals.com

The result for reducible sugar, water solubility and ascorbic acid test for both the fermented and unfermented samples are shown below.

TEST	FERMENTED SAMPLE	UNFERMENTED SAMPLE		
Reducible Sugar	No colour change observed	No colour change observed		
Water Solubility	No reaction at all, it did not dissolve.	No reaction at all, it did not dissolve. Kept going down after each tap and when stirred, it mixed, but after settling for a minute, there were particles below. Therefore it's not totally soluble		
Ascorbic Acid	A very little colour change from blue to pink was observed	From initial light blue to pink colour change was observed		

Table 4: Reducible sugar,	water solu	bility and	ascorbic acid
Tuble II Reducible Sugar,	mater bora	winey and	abcor bie acia

REDUCIBLE SUGAR

Benedict's reagent was added to both separate mixture of samples and then heated up to 75°c and observed for any colour change which will indicate the presence of reducible sugar. In this case, both the fermented and unfermented samples had no colour change, which signifies that there is no reducible sugar present for both sample.

WATER SOLUBILITY

Food substance solubility is the amount of the food substance that dissolves into solution, usually with water as solvent. The presence of lipids, which reduces the water absorption capacity of food, also leads to reduced solubility ^[113]. High solubility of food can show high digestibility of the food which may indicate excellent use for infant formula and food.

From the table above, the fermented sample had no reaction at all, meaning it was completely insoluble, but the unfermented sample can be said to be a bit soluble (not completely soluble). Both samples can be said to possess lipids which makes them insoluble or not completely soluble, making them low digestible and poor use for infant formula and food.

ASCORBIC ACIDS

Ascorbic acid also known as vitamin C is a vitamin found in various food substances and can be used for treatment of scurvy (a deficiency of vitamin C) ^[129]. It also functions as an antioxidant.

The two samples' solution (fermented and unfermented) were added to the light blue solution of diluted dichlorophenolindopheol (DCPIP) one drop at a time to observe for colour change (blue to red or pink), both had colour change.

For the fermented sample, there was a delay in the colour change; a reasonable amount had to be poured before a slight colour change was observed as shown in fig. 61 below, which signifies presence of little ascorbic acid. A few drops of the unfermented sample, changed from blue to pink.

4. CONCLUSION

Rice bran is a promising nutritional and functional food that more attention should be paid to for more research and analysis. More comparison with other food substances should be done.

Fermented rice bran appears to be another part that should be experimented on as well because this research only focused on few analyses in comparison with the unfermented (normal) rice bran.

Based on this research, it can be concluded that from the comparison of result, the unfermented rice bran has more potentials when it comes to food production and pharmaceuticals than the fermented rice bran, apart from the starch content. Further analysis can be investigated to prove otherwise, but from the few done on this research, this is just the conclusion.

Unfermented rice bran posses more protein (especially amino acids), more carbohydrate (especially polysaccharides), which makes it more palatable, better shelf life and good flavour retention than the fermented sample. In other words, the

Vol. 10, Issue 3, pp: (63-86), Month: September - December 2023, Available at: www.noveltyjournals.com

nutritional value of the unfermented sample can be said to be better than the fermented sample. In as much as the fermented sample posses more starch than the unfermented sample, the starch in the unfermented has the ability to absorb water better than the fermented sample that is more of moisture, which makes it more palatable in nutrition. The moisture in the fermented sample also makes it unsuitable for food production and reduces its shelf life, giving room for microbial growth.

Functionally, it can also be said that the unfermented sample is of more value than the fermented sample because of its high flowability, high ascorbic acid and high phenol concentration which can be used in pharmaceuticals. Cholesterol control, multifactorial metabolic disorder (MMD), and inflammatory bowel disease (IBD) can be taken care of with rice bran (unfermented). Tumorigenesis on the other hand can be prevented by fermented rice bran ^[83].

So as earlier stated, unfermented rice bran possess more nutritional and functional value than the fermented rice bran from the few analysis carried out in this research. More research should be done on rice bran as it appears to be a promising functional food.

REFERENCES

- [1] Barron, J. "Black Rice Bran, the Next Super food?". Baseline of Health Foundation. 2010.
- [2] Hui, Y.H. Handbook of vegetable preservation and processing. New York: M. Dekker. p. 180. ISBN 978-0-8247-4301-7. OCLC 52942889. **2004**.
- [3] Sadh, P.K.; Kumar, S.; Chawla, P.; Duhan, J.S. Fermentation: A Boon for Production of Bioactive Compounds by Processing of Food Industries Wastes (By-Products). Molecules, 23, 2560. [CrossRef] [PubMed]. 2018.
- [4] Omarini, A.; Dambolena, J.S.; Lucini, E.; Jaramillo Mejía, S.; Albertó, E.; Zygadlo, J.A. Biotransformation of 1,8cineole by solid state fermentation of spent Eucalyptus wastes from the essential oil industry using Pleurotusostreatus and Favolustenuiculus. Folia Microbiol. 61, 149–157. [CrossRef] [PubMed]. 2016.
- [5] Fonseca, G.G.; Avila G.E.; Fossati, S.L.; Antunes, C.A.P.; Vieira, C.J.A. Protein enrichment and digestibility of soft rush (Juncuseffusus) and rice residues using edible mushrooms Pleurotusostreatus and Pleurotussajor-caju. World J. Microbiol. Biotechnol. 25, 449–456. [CrossRef]. 2009.
- [6] Gul,K.; Yousuf,B.; Singh,A.K.; Singh,P.; Wani, A.A. Ricebran: Nutritional values and its emerging potential for development of functional food-A review. Bioact. Carbohyd. Dietary Fibre, 6, 24–30. [CrossRef]. 2015.
- [7] Alauddin, M.; Islam, J.; Shirakawa, H.; Koseki, T.; ArdiansyahandKomai, M.; Waisundara, V., Shiom, N., Eds.; IntechOpen. Rice Bran as a Functional Food: An Overview of the Conversion of Rice Bran into a Super food/Functional Food. In Super food and Functional Food—An Overview of Their Processing and Utilization; London, UK, 2017.
- [8] Alejandra, B.O.; Diana, L.; María, P.Z.; Romina, P.; Marcelo, F.L.; Damián, B.; Julio, A.Z. Upgrading the Nutritional Value of Rice Bran by Solid-State Fermentation with Pleurotussapidus. Pp 2. 2019
- [9] Lemos, M.R.B.; Souza-Soares, L.A. Rice and its byproducts in southern Brazil. J. Exact Sci. Eng. 10, 21–36. 2000.
- [10] https://www.webmd.com/vitamins/ai/ingredientmono-852/rice-bran
- [11] https://microbenotes.com/submerged-fermentation/
- [12] Manpreet, S.; Sawraj, S.; Sachin, D.; Pankaj, S.; Banerjee, U.C. Influence of process parameters on the production of metabolites in solid-state fermentation. Malaysian Journal of Microbiology, 1(2), 1–9. 2005.
- [13] Kapilan, R. Solid state fermentation for microbial products: A review. Archives of Applied Science Research, 7(8), 21–25. **2015**.
- [14] Pandey, A.; Soccol, C.R.; Leo, J.A.R.; Nigam, P. Solid-state fermentation in Biotechnology (pp. 221–230). New Delhi, India: Asiatech Publishers. 2001.
- [15] Earlia, N.; Muslem, R.S; Amin, M.C.R.S.; Prakoeswa, K.; Rinaldi I. The Scientific World Journal, 1, DOI:10.1155/ 2019/8605743. 2019.

- [16] Holker, U;, Hofer, M.; Lenz, J. Biotechnological advantages of laboratoryscale solid-state fermentation with fungi. Applied Microbiology and Biotechnology, 64, 175–186. doi:10.100700253-003-1504-3 PMID:14963614. **2004**.
- [17] Durand, A. Bioreactor designs for solid state fermentation. Biochemical Engineering Journal, 13(2-3), 113–125. doi:10.1016/S1369-703X(02)00124-9. 2003.
- [18] Singhania, R.R.; Patel, A.K.; Soccol, C.R.; Pandey, A. Recent advances in solid-state fermentation. Biochemical Engineering Journal, 44(1), 13–18. doi:10.1016/j.bej.2008.10.019. 2009.
- [19] Sabu, A.; Augur, C.; Swati, C.; Pandey, A. Tannase production by Lactobacillus sp. ASR-S1 under solid-state fermentation, Process. Bioche, 41, 575–580. 2006.
- [20] Urvashi, S.; Pinki, S.; Zoomi, S. Solid State Fermentation: A Novel Approach in Food Processing Technology Using Food Industry Waste. Published 08 December. doi:10.4018/978-7998-1924-0. Ch010. 2019.
- [21] Kapilan, R. Solid state fermentation for microbial products: A review. Archives of Applied Science Research, 7(8), 21–25. **2015**.
- [22] Massarolo, K.C.; Denardi de Souza, T.; Christ R.A.; Badiale F.E.; Almeida de Souza-Soares, L. Influence of cultivation *Rhizopusoryzae*on rice bran on lipid fraction: Fatty acids and phospholipids. *Biocatal. Agric. Biotechnol.* 8, 204–208. [CrossRef]. 2016.
- [23] Oliveira, M.D.S.; Feddern, V.; Kupsk, L.; Cipolatti, E.P.; Badiale-Furlong, E.; Souza-Soares, L.A. Changes in lipid, fatty acids and phospholipids composition of whole rice bran after solid-state fungal fermentation. *Bioresour. Technol.* 102, 8335–8338. [CrossRef]. 2011.
- [24] Jang, H.D.; Yang, S.S. Polyunsaturated fatty acids production with a solid-state column reactor. *Bioresour. Technol.* 99, 6181–6189. [CrossRef]. 2008.
- [25] Mottillo, S.; Filion, K.B.; Genest, J.; Joseph, L.; Pilote, L.; Poirier, P.; Rinfret; Schiffrin, S.E.L.; Eisenberg, M.J. The metabolic syndrome and cardiovascular risk: a systematic review and meta-analysis. J. Am. Coll. Cardiol. 56 (14), 1113–1132. 2010.
- [26] Mohamed, S. Functional foods against metabolic syndrome (obesity, diabetes, hypertension and dyslipidemia) and cardiovasular disease. Trends Food Sci. Technol.35 (2), 114–128. 2014.
- [27] Abdul-Hamid, A.; Luan, Y.S. Functional properties of dietary fibre prepared from defatted rice bran. Food Chem. 68 (1), 15–19. **2000**.
- [28] Quiro's-Sauceda, A.E., Palafox-Carlos, H., Sa'yago-Ayerdi, S.G., Ayala-Zavala, J.F.; BelloPerez, L.A.; Alvarez-Parrilla, E.; De La Rosa, L.A.; Gonza'lez-Co'rdova, A.F.; Gonzalez-Aguilar, G.A. Dietary fiber and phenolic compounds as functional ingredients: interaction and possible effect after ingestion. Food Funct. 5 (6), 1063–1072. 2014.
- [29] Kahlon, T.S.; Chow, F.I.; Sayre, R.N.; Betschart, A.A. Cholesterol-lowering in hamsters fed rice bran at various levels, defatted rice bran and rice bran oil. J. Nutr. 122 (3), 513–519. 1992.
- [30] Nicolosi, R.J.; Austrian, L.M.; Hegsted, D.M. Rice bran oil lowers serum total and low density lipoprotein cholesterol and apo B levels in nonhuman primates. Atherosclerosis 88 (2–3),133–142. **1991**.
- [31] Palou, M.; Sa'nchez, J.; Garc'ıa-Carrizo, F.; Palou, A.; Pico', C. Pectin supplementation in rats mitigates age-related impairment in insulin and leptin sensitivity independently of reducing food intake. Mol. Nutr. Food Res. 59 (10), 2022–2033. 2015.
- [32] Wang, O.; Liu, J.; Cheng, Q.; Guo, X.; Wang, Y.; Zhao, L.; Zhou, F.; Ji, B. Effects of ferulic acid andγ-oryzanol on high-fat and high-fructose diet-induced metabolic syndrome in rats. PLoS One 10(2), e0118135. 2015.
- [33] Ardiansyah; Shirakawa, H.; Shimeno, T.; Koseki, T.; Shiono, Y.; Murayama, T.; Hatakeyama, E.; Komai, M. Adenosine, an identified active component from the Driselase-treated fraction of rice bran, is effective at improving metabolic syndrome in stroke-prone spontaneously hypertensive rats. J. Agric. Food Chem. 57 (6), 2558–2564. 2009.

- [34] Boonloh, K.; Kukongviriyapan, V.; Kongyingyoes, B.; Kukongviriyapan, U.; Thawornchinsombut, S.; Pannangpetch, P. Rice bran protein hydrolysates improve insulin resistance and decrease pro-inflammatory cytokine gene expression in rats fed a high carbohydrate-high fat diet. Nutrients 7 (8), 6313–6329. 2015.
- [35] Ryan, E.P.; Heuberger, A.L.; Weir, T.L.; Barnett, B.; Broeckling, C.D.; Prenni, J.E. Rice bran fermented with Saccharomyces boulardii generates novel metabolite profiles with bioactivity. J. Agric. Food Chem. 59 (5), 1862– 1870. 2011.
- [36] Hu, C.; Zawistowski, J.; Ling, W.; Kitts, D.D. Black rice (Oryzasativa L. indica) pigmented fraction suppresses both reactive oxygen species and nitric oxide in chemical and biological model systems. J. Agric. Food Chem. 51 (18), 5271–5277. 2003.
- [37] Nakornriab, M.; Sriseadka, T.; Wongpornchai, S. Quantification of carotenoid and flavonoid components in brans of some thai black rice cultivars using supercritical fluid extraction and high-performance liquid chromatography-mass spectrometry. J. Food Lipids 15 (4), 488–503. 2008.
- [38] Min, B.; Chen, M.H.; Green, B.W. Antioxidant activities of purple rice bran extract and its effect on the quality of low-NaCl, phosphate-free patties made from channel catfish(Ictalurus punctatus)belly flap meat. J. Food Sci.74(3). 2009.
- [39] Saenjum, C.; Chaiyasut, C.; Chansakaow, S.; Suttajit, M.; Sirithunyalug, B. Antioxidant and anti-inflammatory activities of gamma-oryzanol rich extracts from Thai purple rice bran. J. Med. Plant Res. 6 (6), 1070–1077. 2012.
- [40] Seril, D.N.; Liao, J.; Yang, G.Y.; Yang, C.S. Oxidative stress and ulcerative colitisassociated carcinogenesis: studies in humans and animal models. Carcinogenesis 24 (3), 353–362. 2003.
- [41] Da Silva, B.C.; Lyra, A.C.; Rocha, R.; Santana, G.O. Epidemiology, demographic characteristics and prognostic predictors of ulcerative colitis. World J. Gastroenterol.: WJG 20 (28), 9458. 2014.
- [42] Ritchie, L.E.; Taddeo, S.S.; Weeks, B.R.; Carroll, R.J.; Dykes, L.; Rooney, L.W.; Turner, N.D. Impact of novel sorghum bran diets on DSS-induced colitis. Nutrients 9 (4), 330. 2017.
- [43] Kondo, S.; Kuda, T.; Nemoto, M.; Usami, Y.; Takahashi, H.; Kimura, B. Protective effects of rice bran fermented by Saccharomyces cerevisiae Misaki-1 and Lactobacillus plantarum Sanriki-SU8 in dextran sodium sulphate-induced inflammatory bowel disease model mice. Food Biosci. 16, 44–49. 2016.
- [44] Ruemmele, F.M. Role of diet in inflammatory bowel disease. Ann. Nutr. Metab. 68 (Suppl. 1), 32-41. 2016.
- [45] Kataoka, K.; Ogasa, S.; Kuwahara, T.; Bando, Y.; Hagiwara, M.; Arimochi, H.; Nakanishi, S., Iwasaki, T.; Ohnishi, Y. Inhibitory effects of fermented brown rice on induction of acute colitis by dextran sulfate sodium in rats. Dig. Dis. Sci. 53 (6), 1601–1608. 2008.
- [46] Ren, T.; Tian, T.; Feng, X.; Ye, S.; Wang, H.; Wu, W.; Qiu, Y.; Yu, C.; He, Y.; Zeng, J.; Cen, J. An adenosine A3 receptor agonist inhibits DSS-induced colitis in mice through modulation of the NF-κBsignaling pathway. Sci. Rep. 5, 9047. 2015.
- [47] Kim, C.J.; Kovacs-Nolan, J.A.; Yang, C.; Archbold, T.; Fan, M.Z.; Mine, Y. L-tryptophan exhibits therapeutic function in a porcine model of dextran sodium sulfate(DSS)-induced colitis. J. Nutr. Biochem. 21 (6),468–475. 2010.
- [48] Komiyama, Y.; Andoh, A.; Fujiwara, D.; Ohmae, H.; Araki, Y.; Fujiyama, Y.; Mitsuyama, K.; Kanauchi, O. New prebiotics from rice bran ameliorate inflammation in murine colitis models through the modulation of intestinal homeostasis and the mucosal immune system. Scand. J. Gastroenterol. 46 (1), 40–52. 2011.
- [49] Kim, K.M.; Yu, K.W.; Kang, D.H.; Suh, H.J. Anti-stress and anti-fatigue effect of fermented rice bran. Phytother. Res. 16 (7), 700–702. 2002.
- [50] Jung, E.H.; Ran Kim, S.; Hwang, I.K.; Youl Ha, T. Hypoglycemic effects of a phenolic acid fraction of rice bran and ferulic acid in C57BL/KsJ-db/db mice. J. Agric. Food Chem. 55 (24), 9800–9804. 2007.

- [51] Ardiansyah; Shirakawa, H.; Koseki, T.; Ohinata, K.; Hashizume, K.; Komai, M. Rice bran fractions improve blood pressure, lipid profile, and glucose metabolism in strokeprone spontaneously hypertensive rats. J. Agric. Food Chem. 54 (5), 1914–1920. 2006.
- [52] Ardiansyah; Shirakawa, H.; Koseki, T.; Hashizume, K.; Komai, M. The Driselasetreated fraction of rice bran is a more effective dietary factor to improve hypertension, glucose and lipid metabolism in stroke-prone spontaneously hypertensive rats compared to ferulic acid. Br. J. Nutr. 97, 67–76. 2007.
- [53] Alauddin, M.; Shirakawa, H.; Koseki, T.; Kijima, N.; Budijanto, S.; Islam, J.; Goto, T.; Komai, M. Fermented rice bran supplementation mitigates metabolic syndrome in stroke-prone spontaneously hypertensive rats. BMC Complement. Altern. Med. 16 (1), 442. 2016.
- [54] Tazawa, H.; Okada, F.; Kobayashi, T.; Tada, M.; Mori, Y.; Une, Y.; Sendo, F.;Kobayashi, M.; Hosokawa, M. Infiltration of neutrophils is required for acquisition of metastatic phenotype of benign murine fibrosarcoma cells: implication of inflammation-associated carcinogenesis and tumor progression. Am. J. Pathol. 163 (6), 2221–2232. 2003.
- [55] Onuma, K.; Kanda, Y.; Suzuki-Ikeda, S.; Sakaki, R.; Nonomura, T.; Kobayashi, M.; Osaki, M.; Shikanai, M.; Kobayashi, H.; Okada, F. Fermented brown rice and rice bran with aspergillusoryzae (FBRA) prevents inflammationrelated carcinogenesis in mice, through inhibition of inflammatory cell infiltration. Nutrients 7 (12), 10237–10250. 2015.
- [56] Islam, M.S.; Murata, T.; Fujisawa, M.; Nagasaka, R.; Ushio, H.; Bari, A.M.; Hori, M.; Ozaki, H. Anti-inflammatory effects of phytosterylferulates in colitis induced by dextran sulphate sodium in mice. Br. J. Pharmacol. 154 (4), 812– 824. 2008.
- [57] Mahla, RS; et al. "NIX-mediated mitophagy regulate metabolic reprogramming in phagotic cells during mycobacterial infection". Tuberculosis. 126: 102046. doi:10.1016/j.tube.2020.202046. PMID 33421909. S2CID 231437641. 2021.
- [58] "Regeneration Medicine Partnership in Education". Archived from the original on 25 April. Retrieved 7 May 2015.
- [59] Ferguson-Smith A.C.; Chen Y.F.; Newman M.S.; May L.T.; Sehgal P.B.; Ruddle F.H. "Regional localization of the interferon-beta 2/B-cell stimulatory factor 2/hepatocyte stimulating factor gene to human chromosome 7p15-p21". Genomics. 2 (3): 203-8. doi:10.1016/0888-7543(88)90003-1. PMID 3294161. **1988**.
- [60] Auron P.E.; Webb A.C.; Rosenwasser L.J.; MucciS.f.; Rich A.; Wolff S.M.; Dinarello C.A. "Nucleotide sequence of human monocyte interleukin 1 precursor cDNA". Proceedings of the National Academy of Sciences of the United States of America. 81 (24): 7907-11. Bibcode:1984PNAS...81.7907A. doi:10.1073/pnas.81.24.7907. PMC 392262. PMID 6083565. 1984.
- [61] March C.J.; Mosley B.; Larsen A.; Cerreti D.P.; Braedt G.; Price V.; et al. "Catabolin" is the name given by Jeremy Saklatvala for IL-1 alpha. "Cloning, sequence and expression of two distinct human interleukin-1 complementary DNAs". Nature. 315 (6021): 641-7. Bibcode:1985Natur.315..641M. doi:10.1038/315641a0. PMID 2989698. S2CID 4240002. 1985.
- [62] Clark B.D.; Collins K.L.; Gandy M.S.; Webb A.C.; Auron P.E. "Genomic sequence for human prointerleukin 1 beta: possible evolution from a reverse transcribed prointerleukin 1 alpha gene". Nucleic Acids Research. 14 (20): 7897-914. Doi:10.1093/nar/14.20.7897. PMC 311823. PMID 3490654. 1986.
- [63] Bensi G.; Raugei G.; Palla E.; Carinci V.; Tornese B.D.; Melli M. "Human interleukin-1 beta gene". Gene. 52 (1): 95-101. doi:10.1016/0378-1119(87)90398-2. PMID 2954882. 1987.
- [64] Lloyd, B.J.; Siebenmorgen, T.J.; Beers, K.W. Effects of commercial processing on antioxidants in rice bran. Cereal Chem. 77 (5), 551–555. 2000.
- [65] Rao, M.K.; Achaya, K.T. Unsaturated fatty acids as synergists for antioxidants. Eur. J. Lipid Sci. Technol. 70 (4), 231–234. **1968**.

- [66] Richard, D.; Kefi, K.; Barbe, U.; Bausero, P.; Visioli, F. Polyunsaturated fatty acids as antioxidants. Pharmacol. Res. 57 (6), 451–455. 2008.
- [67] Faria, S.A.D.S.C.; Bassinello, P.Z.; Penteado, M.D.V.C. Nutritionalcompositionofricebran submitted to different stabilization procedures. Braz. J. Pharm. Sci. 48 (4), 651–657. 2012.
- [68] Bhosale, S.; Vijayalakshmi, D. Processing and nutritional composition of rice bran. Curr. Res. Nutr. Food Sci. J. 3 (1), 74–80. 2015.
- [69] Zubair, M.A.; Rahman, M.S.; Islam, M.S.; Abedin, M.Z.; Sikder, M.A. A comparative study of the proximate composition of selected Rice varieties in Tangail, Bangladesh. J. Environ. Sci. Nat. Resour. 8 (2), 97–102. 2016.
- [70] Tuncel, N.B.;Yılmaz, N. Gamma-oryzanol content, phenolic acid profiles and antioxidant activity of rice milling fractions. Eur. Food Res. Technol. 233 (4), 577. 2011.
- [71] Rogers, E.J.; Rice, S.M.; Nicolosi, R.J.; Carpenter, D.R.; McClelland, C.A.; Romanczyk, L.J. Identification and quantitation ofγ-oryzanol components and simultaneous assessment of tocols in rice bran oil. J. Am. Oil Chem. Soc. 70 (3), 301–307. 1993.
- [72] Terahara, N.; Saigusa, N.; Ohba, R.; Ueda, S. Composition of anthocyanin pigments in aromatic red rice and its wine. Nippon Shokuhin Kogyo Gakkaishi 41 (7), 519–522. 1994.
- [73] Harukaze, A.; Murata, M.; Homma, S. Analyses of free and bound phenolics in rice. Food Sci. Technol. Res. 5 (1), 74–79. 1999.
- [74] Arai, S.; Osawa, T.; Ohigashi, H.; Yoshikawa, M.; Kaminogawa, S.; Watanabe, M.; Ogawa, T.; Okubo, K.; Watanabe, S.; Nishino, H.; Shinohara, K. A mainstay of functional food science in Japan—history, present status, and future outlook. Biosci. Biotechnol. Biochem. 65 (1),1–13. 2001.
- [75] Saito, M. Role of FOSHU (food for specified health uses) for healthier life. Yakugakuzasshi: J. Pharm. Soc. Jpn. 127 (3), 407–416. 2007.
- [76] Thomas, P.R.; Earl, R. Opportunities in the Nutrition and Food Science: Research Challenges and the Next Generation of Investigators. Committee on Opportunities in the Nutrition and Food Sciences, Food and Nutrition Board, Institute of Medicine Enhancing the Food Supply. 98-142 National Academy Press Washington, DC. 1994.
- [77] American Dietetic Association (ADA). Position of the American dietetic association and dietitians of Canada: vegetarian diets. J. Acad. Nutr. Diet. 103 (6), 748. 2003.
- [78] Hasler, C.M. Functional foods: benefits, concerns and challenges—a position paper from the American Council on Science and Health. J. Nutr. 132 (12), 3772–3781. 2002.
- [79] Martirosyan, D.M.; Singh, J. A new definition of functional food by FFC: what makes a new definition unique? Funct. Foods Health Dis. 5 (6), 209–223. 2015.
- [80] Krikorian, R.; Nash, T.A.; Shidler, M.D.; Shukitt-Hale, B.; Joseph, J.A. Concord grape juice supplementation improves memory function in older adults with mild cognitive impairment. Br. J. Nutr. 103 (5), 730–734. **2010**.
- [81] Kuno, T.; Nagano, A.; Mori, Y.; Kato, H.; Nagayasu, Y.; Naiki-Ito, A.; Suzuki, S.; Mori, H.; Takahashi, S. Preventive effects of fermented brown rice and rice bran against prostate carcinogenesis in TRAP rats. Nutrients 8 (7), 421. 2016.
- [82] Sharma, R.; Srivastava, T.; Saxena, D.C. International Journal of Engineering Research and Applications, 5,107. 2015.
- [83] Amarasinghe, B.M.; Kumarasiri, W.P.K.; M.P.M.; Gangodavilage, N.C. Food and Bioproducts Processing, 87,108, DOI:10.1016/j.fbp.2008.08.002. 2009.
- [84] Yilmaz, N. Food Chemistry, 190, 179(2016), DOI:10.1016/j. foodchem. 2015.05.094. 2016.

- [85] Patil, S.S.; Kar, A.; Mohapatra, D. Food and Bioproducts Processing, 99, 204(2016), DOI:10.1016/j.fbp. 2016.05.002. 2016.
- [86] Patel, M.; Naik, S.N.; Journal of Scientific and Industrial Research, 63, 569. 2004.
- [87] Prabhakar, J.V.; Venkatesh, K.V.L. Chemistry and Materials Science, 63, 644, DOI:10.1007/BF02638229. 1986.
- [88] Spark, D.; Hernandez, R.; Zappi, M.; Blackwell, D.; Fleming, T. Journal of the American Oil Chemists' Society, 83, 10, DOI:10.1007/s11746-006-5042-x. 2006.
- [89] Kong, W.; Kang, Q.; Feng, W.; Tan, T. Chemical Engineering Research and Design, 104, 1, DOI:10.1016/ j.cherd.2015.06.001. 2015.
- [90] Proctor, A.; Bowen, D.J. Journal of the American Oil Chemists' Society, 73, 811, DOI:10.1007/BF02517960. 1996.
- [91] Mohd-Daud, N.S.; Zaidel, D.N.A.; Lai, K.S.; Khairuddin, N.; Mohd-Jusoh, Y.M.; Muhamad, I.I. Arabian Journal for Science and Engineering, 43, 6237, DOI:10.1007/s13369-018-3438-1. 2018.
- [92] Garba, U.; Singanusong, R.; Jiamyangyuen, S.; Thongsook, T. In Proceedings of the 4th International Conference on Rice Bran Oil, Bangkok, Thailand, pp. 1-12. **2017**.
- [93] Tomita, K.; Machmudah, S.; Wahyudiono; Fukuzato, R.; Quitain, A.T.; Sasaki, M.; Goto, M.; Kanda, H. Separation and Purification Technology, 125, 319, DOI:10.1016/j.seppur.2014.02.008. 2014.
- [94] Khoei, M.; Chekin, F. Food Chemistry, 194, 503, DOI:10.1016/j.foodchem.2015.08.068. 2016.
- [95] Djaeni, M.; Listyadevi, Y.L. Journal of Physics: Conference Series, 1295, 012027, DOI:10.1088/1742-6596/1295/1/012027. 2019.
- [96] Shukla, H.S.; Pratap, A. Journal of Oleo Science. 66, 973, DOI:10.5650/jos.ess17067. 2017.
- [97] Sarah Moore. Why is Moisture Content Analysis of Food Important? 2020
- [98] Iwe, M.O.; Onyeukwu, U.; Agiriga, A.N. Proximate, functional & pasting properties of FARO 44 rice, African yam bean and brown cowpea seeds composite flour.Cogent Food & Agriculture, 2: 1142409. http://dx.doi.org/10.1080/ 23311932.2016.1142409. 2016
- [99] Suresh, C.; Samsher, S.Assessment of functional properties of different flours. African Journal of Agricultural Research.8(38); 4849-4852. DOI:10.5897/AJAR2013.6905. ISSN 1991-637X ©. 2013
- [100] Suresh, C.; Samsher, S.; Durvesh, K. Evaluation of functional properties of composite flours and sensorial attributes of composite flour biscuits. J Food SciTechnol, 52(6):3681–3688. DOI 10.1007/s13197-014-1427-2. 2015
- [101] Jitngarmkusol, S.; Hongsuwankul, J.; Tananuwong, K. Chemical composition, functional properties and microstructure of defatted macademice flours. Food Chem. 110:23–30. 2008
- [102] Zghal, M.C.; Scanlon, M.G.; Sapirstein, H.D. "Effects of Flour Strength, Baking Absorption, and Processing Conditions on the Structure and Mechanical Properties of Bread Crumb." Cereal Chemistry Journal, 78.1: 1-7. 2001
- [103] Puhr, D.H.; D'Appolonia, B.L. "Effect of Baking Absorption on Bread Yield, Crumb Moisture, and Crumb Water Activity." Cereal Chem. 69.5: 582-86. 1992
- [104] Pyler, E.J. "Physical and Chemical Test Methods." Baking Science and Technology. Merriam, Kan.: Sosland Pub. 851-71. 1988
- [105] Serna-Saldivar, S.O. Quality Control of the Cereal Grains and Their Products. Cereal Grains: Properties, Processing, and Nutritional Attributes, CRC Press, Taylor & Francis Group, pp. 491–493. 2010
- [106] Sreerama, Y.N.; Sashikala, V.B.; Pratape, V.M.; Singh, V. Nutrients and antinutrients in cowpea and horse gram flours in comparison to chickpea flour: Evaluation of their flour functionality. Food Chemistry, 131(2), 462–468.
 2012

- Vol. 10, Issue 3, pp: (63-86), Month: September December 2023, Available at: www.noveltyjournals.com
- [107] Kuntz, I.D. Hydration of macromolecules III. Hydration of polypeptides. J Am ChemSocie 93:514-515. 1971
- [108] Butt, M.S.; Batool, R. Nutritional & functional properties of some promising legumes proteins isolates. Pakistan J Nutr, 9(4):373–379. 2010
- [109] Buckman, H.O.; Brady, N.C. Nature & Property of Soils A College Text of Edaphology (Sixth edition), New York: Macmillan Publishers, New York, NY. Page 50. 1960
- [110] The Powder process. The Powder Bulk Density –the Bulk Solids density –the Bulk Powder Properties –the Powder Loose Density –the Powder tapped density PowderProcess.net. *www.powderprocess.net*. **2018**
- [111] Oppong, D.; Eric, A.; Samuel, O.; Eric, B.; Patrick, S. Proximate Composition & Some Functional Properties of Soft Wheat Flour. Intl J. of Innovative Research in Sci., Engineering and Technol, 4 (2): 753-758. DOI: 10.15680/ IJIRSET.2015.0402097. 2015
- [112] IUPAC. Compendium of Chemical Terminology. 2nd ed. (the "Gold Book") (1997). Online corrected version: "Solubility". doi:10.1351/goldbook.S05740. 2006
- [113] Clugston, M.; Fleming, R. Advanced Chemistry (1st ed.). Oxford: Oxford Publishing. p. 108. 2000
- [114] Rogers, E.; Stovall, I. Fundamentals of Chemistry: Solubility. Department of Chemistry. University of Wisconsin.2000
- [115] Pratt, C.W.; Cornely, K. Essential Biochemistry (Third ed.). Wiley. p. 626. ISBN 978-1118083505. 2013
- [116] Nelson, D.L.; Cox, M.M. Lehnniger: Principles of Boichemistry (Fifth ed.). W.H. Freeman and Company. p. 241. ISBN 978-0716771081. 2008
- [117] Dills, W.L.Jr. "Protein fructosylation: fructose and the Maillard reaction". The American Journal of Clinical Nutrition. American Society for Nutrition. 58 (5): 779S – 87. doi:10.1093/ajcn/58.5.779s. ISSN 0002-9165. PMID 8213610. 1993
- [118] Jiang, Z; Wang, L; Wu, W.; Wang, Y. "Biological activities and physiochemical properties of Mailard reaction products in sugar-bovine casein peptide model system". Food Chemistry. Elsevier. 141 (4): 3837-3845. doi:10.1016/ j.foodchem.2013.06.041. ISSN 0308-8146. PMID 23993556. 2013
- [119] Pedreschi, F.; Mariotti, M.S.; Granby, K. "Current issues indietry acrylamide: formation, mitigation and risk assessment". Journal of the Science of Food and Agriculture. Society of Chemistry Industry. 94 (1): 9 20. doi:10. 1002/jsfa.6349. hdi:10533/127076. ISSN 0022-5142. PMID 23939985. 2013
- [120] American Cancer Society. "Acylamide and Cancer Risk". 2019
- [121] Leoterio, D.M.S.; Silva, P.; Souza, G.; Alves. A.A.; Belian, M.; Galembeck, A.; Lavorente, A.F. "Copper-4,4'dipyridyl coordination compound as solid reagent for spectrophotometric determination of reducing sugar employing a multicommutation approach". Food Control. European Federation of Food Science and Technology; International Union of Food Science and Technology. 57: 225 – 231. doi:10.1016/j.foodcont.04.017. ISSN 0956-7135. 2015
- [122] Allen, D.W; Cooksey, C.; Tsai, B.K. "Spectrophotometry". NIST. 2009
- [123] ELGA LabWater (ELGA VEOLIA). "Spectrophotometry" (https://www.elgalabwater.com/applications/spectro photometry). 2021
- [124] Michael, K.S.; George A.Z.; Aristidis, N.A.; Themistoclis, A.K. Spectrophotometric Determination of Phenols and Cyanides After Distillation From Natural Waters. 'Laboratory of Analytical Chemistry and Laboratory of Environmental Pollution Control, Arisiotle University of Thessaloniki, 54006 Thessaloniki, Greece (Received I5 October 1999; In final form 22 March). 2000
- [125] Moore, J.W.; Ramamoorthy, S. Organic Chemicals in Narural Waters (Springer-Verlag, New York) pp. 141 167.
 1984

- Vol. 10, Issue 3, pp: (63-86), Month: September December 2023, Available at: www.noveltyjournals.com
- [126] Waggot, A.; Wheatland, A.B. Proceedings of the 2nd International Symposium on Aquatic Pollutants, Amsterdam, The Netherlands. September. 1977
- [127] The American Society of Health-System Pharmacists. "Ascorbic Acid" (https://www.drugs.com/monograph/ ascorbic-acid.html). Archived (https://web.archive.org/web/20161230161611/https://www.dru gs.com/monograph/ ascorbic-acid.html) from the original on December 30. Retrieved December 8. 2016
- [128] Dietary Reference Intakes for Vitamin C, Vitamin E, Selenium, and Carotenoids. Washington, DC: The National Academies Press. "Vitamin C" (https://www.nap.edu/read/9810/chapter/7). 2000. pp. 95– 185. ISBN 978-0-309-06935-9. Archived (https://web.archive.org/web/20170902180153/https://www.nap.edu/read/9810/chapter/7) from the original on September 2. Retrieved September 1. 2017
- [129] Office of Dietary Supplements, US National Institutes of Health. "Fact Sheet for Health Professionals Vitamin C" (https://ods.od.nih.gov/factsheets/VitaminC-HealthProfessional/). February 11, 2016. Archived (https://web. archive.org/web/20170730052126/https://ods.od.nih.gov/factsheets/VitaminC-HealthProfessional/)from the original on July 30, 2017.
- [130] Stuart, M.C.; Kouimtzi, M.; Hill, S.R. (eds.). WHO Model Formulary 2008. World Health Organization. hdl:10665/44053 (https://hdl.handle.net/10665%2F44053). ISBN 9789241547659. 2009
- [131] Drugs.com. "Ascorbic acid Use During Pregnancy" (https://www.drugs.com/pregnancy/ascorbic-acid.html). Archived (https://web.archive.org/web/20161231075819/https://www.drugs.com/pregnanc y/ascorbic-acid.html) from the original on December 31. Retrieved December 30. 2016
- [132] Corvallis, O.R. "Vitamin C" (http://lpi.oregonstate.edu/mic/vitamins/vitamin-C). Micronutrient Information Center, Linus Pauling Institute, Oregon State University. Retrieved June 19, 2019. July 1, 2018
- [133] Magiorkinis, E.; Beloukas, A.; Diamantis, A. "Scurvy: past, present and future". The European Journal of Internal Medicine. 22 (2): 147–52. doi:10.1016/j.ejim.2010.10.006 (https://doi.org/10.1016% 2Fj.ejim.2010.10.006).
 PMID 21402244 (https://pubmed.ncbi.nlm.nih.gov/21402244). 2011
- [134] Hodges, R.E.; Baker, E.M.; Hood, J.; Sauberlich, H.E.; March, S.C. "Experimental scurvy in man". The American Journal of Clinical Nutrition. 22 (5): 535–48. doi:10.1093/ajcn/22.5.535 (https://doi.org/10.1 093%2Fajcn%2F 22.5.535). PMID 4977512 (https://pubmed.ncbi.nlm.nih.gov/4977512). 1969
- [135] Baron, J.H. "Sailors' scurvy before and after James Lind--a reassessment". Nutrition Reviews. 67 (6): 315–32. doi:10.1111/j.1753-4887.2009.00205.x (https://doi.org/10.1111%2Fj.1753-4887.2009.00205.x). PMID 19519673 (https://pubmed.ncbi.nlm.nih.gov/19519673). S2CID 20435128 (https://api.semanticscholar.org/CorpusID:2043 5128). 2009
- [136] Lind, J. A Treatise of the Scurvy. London: A. Millar. In the 1757 edition of his work, Lind discusses his experiment starting on page 149. (https://archive.org/stream/treatiseonscurvy00lind#page/149/mode/1up) Archived (https://web. archive.org/web/20160320155753/https://archive.org/stre am/treatiseonscurvy00lind) March 20, 2016, at the Wayback Machine. 1753
- [137] Ashor, A.W.; Lara, J.; Mathers, J.C.; Siervo, M. "Effect of vitamin C on endothelial function in health and disease: a systematic review and meta-analysis of randomised controlled trials". Atherosclerosis. 235 (1): 9–20. doi:10.1016/j.atherosclerosis.04.004 (https://doi.org/10.1016%2F j.atherosclerosis.2014.04.004). PMID 24792921 (https://pubmed.ncbi.nlm.nih.gov/24792921). 2014
- [138] Medlineplus.gov. "Vitamin C: MedlinePlus Medical Encyclopedia" (https://web.archive.org/web/20160728013605/ https:// medlineplus.gov/ency/article/002404.htm). Archived from the original (https://medlineplus.gov/ency/article/ 002404.htm) on July 28. Retrieved July 23. 2016
- [139] Pauling, L. Vitamin C, the Common Cold, and the Flu. W.H. Freeman and Company. 1976

- Vol. 10, Issue 3, pp: (63-86), Month: September December 2023, Available at: www.noveltyjournals.com
- [140] Goodwin, J.S.; Tangum, M.R. "Battling quackery: attitudes about micronutrient supplements in American academic medicine". Archives of Internal Medicine. 158 (20): 2187–91. doi:10.1001/archinte.158.20.2187 (https://doi.org/ 10.1001%2Farchinte.158.20.2187). PMID 9818798 (https://pubmed.ncbi.nlm.nih.gov/9818798). 1998
- [141] Naidu, K.A. "Vitamin C in human health and disease is still a mystery? An overview" (http://www.nutritionj.com/content/pdf/1475-2891-2-7.pdf) (PDF). Nutrition Journal. 2 (7): 7. doi:10.1186/1475-2891-2-7 (https://doi.org/10.1 186%2F1475-2891-2-7). PMC 201008 (https://www.ncbi.nlm.nih.gov/pmc/articles/PMC201008). PMID 14498993 (https://pubmed.ncbi.nlm.nih.gov/14498993). Archived (https://web.archive.org/web/20120918153239/http://www.nutritionj.com/content/p df/1475-2891-2-7.pdf) (PDF) from the original on September 18, 2012. 2003
- [142] Thomas, L.D.; Elinder, C.G.; Tiselius, H.G.; Wolk, A.; Akesson, A. "Ascorbic acid supplements and kidney stone incidence among men: a prospective study" (https://doi.org/10.1001%2Fjamainternmed.2013.2296). JAMA Internal Medicine. 173 (5): 386–8. doi:10.1001/jamainternmed.2013.2296 (https://d oi.org/10.1001%2Fjamainternmed. 2013.2296). PMID 23381591 (https://pubmed.ncbi.nlm.nih.gov/23 381591). 2013
- [143] Bowker, M.I.; Heinrich, P.S. "Preparation of Water-Soluble Compounds through Salt Formation." In Camille Georges Wermuth, ed. The Practice of Medical Chemistry, pp. 747–766. Burlington, MA: Elsevier, p. 756. 2008
- [144] Podczeck; Fridun; Brian, E.J. Pharmaceutical Capsules. London: Pharmaceutical Press, p.111. eds. 2007
- [145] "General Chapters: BULK DENSITY AND TAPPED DENSITY" (http://www.pharmacopeia.cn/v29240/usp29 nf24s0_c616.html).
- [146] Kanig, J.L.; Lachman, L.; Lieberman, H.A. The Theory and Practice of Industrial Pharmacy (3 ed.). Philadelphia: Lea & Febiger. ISBN 0-8121-0977-5. 1986
- [147] Beddow, J.K. "Professor Dr. Henry H. Hausner, 1900–1995." Particle & Particle Systems Characterization 12: 213. doi:10.1002/ppsc.19950120411 (https://doi.org/10.1002%2Fppsc.19950120411) .1995
- [148] Grey, R.O.; Beddow, J.K. "On the Hausner Ratio and its relationship to some properties of metal powders" Powder Technology, Vol.2, No.6, pp 323-326. 1969
- [149] Li, Q. et al. "Interparticle van der Waals force in powder flowability and compactibility" International Journal of Pharmaceutics, Vol.280, Iss.1-2, pp 77-93. 2004
- [150] Conesa, C. et al. "Characterization of Flow Properties of Powder Coatings Used in the Automotive Industry" Kona, Vol.22, pp 94-106. 2004
- [151] Rough, S.L.; Wilson, D.I.; York, D.W. "Effect of solids formulation on the manufacture of high shear mixer agglomerates" Adv. Powder Technol., Vol.16, pp 145-169. 2005
- [152] Garcia, R.A.; Flores, R.A.; Mazenko, C.E. "Factors contributing to the poor bulk behavior of meat and bone meal and methods for improving these behaviors" Bioresource Technology, Vol.98, No.15, pp 2852-2858. 2007
- [153] . USP <1174> "Excipient General Information Chapter: Powder Flow
- [154] Cain, J. "An alternative technique for determining ANSI/CEMA standard 550 flowability ratings for granular materials" Powder Hand. Proc, Vol.14, No.3, pp 218-220. 2002
- [155] Awuchi, C.G.; Igwe, V.; Echeta, C.K. The Functional Properties of Foods and Flours. https://www.researchgate.net/ publication/337403804. International Journal of Advanced Academic Research | Sciences, Technology and Engineering | ISSN: 2488-9849 Vol. 5, Issue 11 November 2019
- [156] Beth Israel Lahey Health (Winchester Hospital). "Gamma Oryzanol".